

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property
Organization
International Bureau



(43) International Publication Date
2 September 2004 (02.09.2004)

PCT

(10) International Publication Number
WO 2004/073670 A1

(51) International Patent Classification²: A61K 7/16 (81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, YU, ZA, ZM, ZW.

(21) International Application Number: PCT/GB2004/000515

(22) International Filing Date: 11 February 2004 (11.02.2004)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
0303676.1 18 February 2003 (18.02.2003) GB

(71) Applicant (for all designated States except US): QUEST INTERNATIONAL SERVICES B.V. [NL/NL]; Huiserstraatweg 28, NL-1411 GP Naarden (NL).

(72) Inventors; and

(75) Inventors/Applicants (for US only): BEHAN, John, Martin [GB/GB]; The Forge, The Green, Boughton Aluph, Kent TN25 4JB (GB). BRADSHAW, David, Jonathan [GB/GB]; 10 Rookery Close, Kennington, Ashford, Kent, TN24 9RP (GB). RICHARDS, Jonathan [GB/GB]; 153 Canterbury Road, Kennington, Ashford, Kent TN24 9QB (GB). MUNROE, Michael, John [GB/GB]; The Oast House, Lacton Oast, Willesborough, Ashford, Kent TN24 0JF (GB).

(74) Agent: KEITH W NASH & Co; 90-92 Regent Street, Cambridge CB2 1DP (GB).

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HU, IE, IT, LU, MC, NL, PT, RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Declaration under Rule 4.17:

— of inventorship (Rule 4.17(iv)) for US only

Published:

— with international search report
— before the expiration of the time limit for amending the claims and to be republished in the event of receipt of amendments

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

(54) Title: IMPROVEMENTS IN OR RELATING TO FLAVOUR COMPOSITIONS

(57) Abstract: A flavour composition comprising at least two flavour materials selected from the following Group (a) materials: decanol, nonanol, decanal, anethole synthetic, cardamom oil, cinnamic aldehyde, ionone alpha, origanum, tarragon, thymol; and at least one flavour material selected from the following Group (b) materials: nonanal, Aniseed rectified, basil oil, camomile oil, citral, clove bud oil, Damascene E, ginger, Tea Tree Pure, peppermint oil of natural origin. The compositions may reduce or prevent dental caries.

WO 2004/073670 A1

Title: Improvements in or relating to flavour compositions

Field of the Invention

This invention relates to flavour compositions, i.e. a mixture of flavour materials, to products containing such flavour compositions, and to the use of a flavour material or flavour composition for inhibiting or reducing acid-producing oral bacteria, particularly the bacterium *Streptococcus mutans*, implicated in initiating dental caries (tooth decay).

Background to the Invention

Dental plaque is a soft whitish material which forms on the surfaces of the teeth. Plaque is a matrix of bacteria, bacterial products and salivary and other host-derived components. One of the bacteria present in dental plaque is *Streptococcus mutans*, which converts dietary sugar into dextran, an insoluble, inert gelatinous polysaccharide which enables the bacterium to adhere to the tooth surface.

The excessive and/or frequent consumption of fermentable dietary sugars can lead to the enrichment of particular groups of bacteria, especially *Streptococcus mutans*, in dental plaque. Dental caries results from the dissolution of tooth enamel (demineralisation) by organic acids generated by bacteria. *Streptococcus mutans* is particularly important in the formation of dental caries because the bacterium rapidly generates large quantities of lactic acid from dietary sugars, whilst concomitantly displaying unusual acid tolerance and tenacity to tooth surfaces.

A number of strategies are employed to combat the development of dental caries.

One approach includes mechanical oral hygiene measures, e.g. brushing, to physically remove plaque. However, brushing alone is insufficient to remove all plaque that may form on the teeth or to prevent the formation of further plaque.

Numerous oral care compositions are known which include a variety of antimicrobial compounds, such as sodium dodecyl sulphate, essential oils, and other miscellaneous agents for inhibiting the development of dental caries. For example, WO 98/44901 concerns oral hygiene compositions including an antimicrobial agent selected from cedarwood oil, chloramphenicol, citronella oil, *Glycyrrhiza glabra* extract, juicy fruit basil oil, lemon basil oil, and *Rosmarinus officinalis* oil; EP 0819380 describes p-oxybenzoic acid esters such as methyl, ethyl and butyl p-oxybenzoates which possess bacteriocidal, antibacterial or bacteriostatic activity against *Streptococcus mutans*; US 4661342 describes oral compositions containing hydroxamic acids such as 2-(4-butoxyphenyl)acetohydroxamic acid to prevent *Streptococcus mutans* from colonising in the oral cavity; US 4590215 discloses that 1-alpha-cadinol inhibits *Streptococcus mutans* growth, polyunsaturated long-chain alcohols such as linolenyl alcohol and linoleyl alcohol are also described for use to this effect in US 4372978; DE 4221103 discloses mixtures of myrrh extract or oil, mulberry bark extract, *Cimicifuga heraleifolia* extract and green tea extract, as having antibacterial activity against cariogenic bacteria. A number of these antimicrobial strategies will also inhibit the generation of acid from fermentable dietary sugars.

A further strategy for combating the development of dental caries is the replacement of conventional, readily fermentable dietary sugars in consumable products with non-fermentable ingredients, e.g. sweeteners, weakly fermentable sugars, or sugars, which interfere with normal sugar metabolism. For example, US 5294449 discloses the use of erythrose in chewing gum to deliver anti-caries properties; GB 2046757 discloses the use of aldosylfructoside in this way; and EP 0438912 describes an edible composition which includes a bulking agent comprising polydextrose, an encapsulated flavouring agent and an effective amount of an intense sweetening agent.

Still other approaches interfere with the formation of the polysaccharide dextran in order to reduce the adherence of cariogenic bacteria to teeth or plaque. For example, EP 0704202 discloses cycloisomaltooligosaccharide as an active which inhibits glucan synthetase from *Streptococcus mutans*; and US 4912089 discloses inhibition of glucan production by *Streptococcus mutans* using a purified Gymnemic acid derived from *Gymnema sylvestre*.

One of the most important caries preventive measures known is the use of fluoride which may be delivered, e.g. via consumer products such as oral care products or via fluoridation of public drinking water supplies. Drinking water is usually fluoridated using sodium fluoride, whereas, in the case of oral care products, fluoride may be incorporated in the form of a variety of salts including sodium salts, e.g. sodium fluoride and sodium monofluorophosphate, strontium salts, calcium salts etc. Fluoride typically acts to prevent caries by its incorporation into dental enamel. Such incorporation renders dental enamel less susceptible to demineralisation, whilst also promoting remineralisation. Fluoride is also known to inhibit a variety of bacterial metabolic processes, in particular, carbohydrate metabolism. Several documents also disclose the combination of fluoride with other agents. For example, US 2002068039 discloses the use of a grapefruit seed extract in synergistic combination with a fluoride ion-providing compound to inhibit the growth and metabolism of, and to kill, plaque bacteria.

Summary of the Invention

The present invention is based on extensive testing of flavour materials to determine whether a particular material is capable of inhibiting the production of acid from the metabolism of dietary sugar by micro-organisms present in the oral cavity, and more particularly inhibiting lactic acid production from glucose by *Streptococcus mutans*. Based on this testing, flavour materials were identified, which whilst known, may possess hitherto unappreciated properties in terms of inhibiting or reducing acid-producing bacteria. The invention thus enables flavour compositions to be defined that reduce or prevent dental caries. Additionally, in a preferred embodiment, the invention enables flavour compositions to be formulated comprising flavour material(s) which selectively

target and inactivate the acid-producing bacteria whilst preserving the remaining protective oral cavity microflora.

Accordingly, in one aspect, the present invention provides a flavour composition comprising at least two flavour materials selected from the following Group (a) materials: decanol, nonanol, decanal, anethole synthetic, cardamom oil, cinnamic aldehyde, ionone alpha, origanum, tarragon, thymol; and at least one flavour material selected from the following Group (b) materials: nonanal, Aniseed rectified, basil oil, camomile oil, citral, clove bud oil, Damascone F, ginger, Tea Tree Pure, peppermint oil of natural origin.

Cardamom oil is conveniently cardamom English.

Cinnamic aldehyde is conveniently cinnamic aldehyde extra, available from Quest International.

Basil oil is conveniently basil comores.

Camomile oil is conveniently camomile English.

Clove bud oil is preferably rectified, e.g. clove bud rectified extra.

For enhanced inhibition of acid-producing bacteria, preferably, flavour compositions of the invention comprise at least three flavour materials from Group (a).

Also preferred are flavour compositions comprising at least two flavour materials from Group (b).

Flavour compositions in accordance with the invention preferably comprise at least 3% by weight, more preferably at least 6% by weight and even more preferably at least 10% by weight, of flavour materials from Group (a); and preferably at least 3% by weight, more

preferably at least 10% by weight and even more preferably at least 25% by weight, of flavour materials from Group (b).

Conveniently, Group (a) and Group (b) flavour materials may together comprise at least 6% by weight of the total weight of the flavour composition, preferably at least 15% by weight, more preferably at least 30% by weight, even more preferably at least 40% by weight and most preferably at least 50% by weight.

Peppermint oil useful herein is of natural origin. Preferably, the peppermint oil is a Piperita type from the far west regions of the United States, e.g. Peppermint American Rectified, Peppermint American Yakima Rectified, Peppermint American Willamette Natural, which is preferably rectified. Also preferred for use in a composition of the invention is an Arvensis type peppermint oil, typically having a total terpene content of less than 3.2% by weight, e.g. Peppermint Indian Rectified, Peppermint Arvensis Terpeneless ACF153, Peppermint Chinese Triple Rectified (available from Quest International).

The ingredients of the composition are known flavour materials which are readily available commercially in grades suitable for various intended purposes. Details of the flavour materials and potential suppliers thereof are mentioned, for example, in "Allured's Flavor and Fragrance Materials 2002", Allured Publishing Corp., Carol Stream, Illinois, USA, ISBN 0-931710-84-7.

Also included within the scope of the invention is a method, particularly a cosmetic method, for reducing or preventing acid-producing bacteria by introducing in the oral cavity a flavour composition in accordance with the invention.

The flavour materials useful in a flavour composition of the invention are capable of inhibiting the production of acid by micro-organisms present in the oral cavity. In

particular, the flavour materials are capable of inhibiting the production of lactic acid from glucose by the bacterium *Streptococcus mutans* present in the oral cavity.

One property that characterises the effectiveness of a compound, e.g. a flavour material, to inhibit the production of acid by the micro-organism *Streptococcus mutans* in the oral cavity, is the minimum inhibitory concentration, or MIC, of the compound. The MIC is the minimum amount of a compound (e.g. in ppm) at which no bacterial growth is observed. Generally, the lower the MIC of a compound for a bacterium, the more effective the compound will be at inhibiting bacterial growth. At concentrations above the MIC, a compound may act by directly killing existing viable bacteria or inhibiting the growth and reproduction of the bacteria (antimicrobial effect). At concentrations below the MIC, a compound may interfere with the metabolic process, e.g. by inactivating the bacteria producing acid, but typically does not inhibit the growth and reproduction of bacteria (sub-lethal or sub-MIC effect).

The inhibitory effect of a flavour composition comprising the flavour materials useful herein can be achieved antimicrobially, or more surprisingly, sub-lethally.

The antimicrobial effects of compounds, e.g. flavour materials, are usually divided into two types; they can either inhibit bacterial growth (bacteriostatic action) or alternatively they can act by directly killing existing viable bacteria (bactericidal action).

The bacteriostatic action of a compound "X" (such as a flavour material) against a particular bacterium, can be tested for *in vitro* by inoculating a standard, small number of bacteria into broths containing an appropriate range of concentrations of X. The broths are then incubated for a suitable time, and growth compared with a control containing no inhibitor. The broth containing the lowest concentration of X which shows reduction of growth compared to the control broth, is defined as the minimum inhibitory concentration (MIC).

The determination of the bactericidal action of a compound "Y" (such as a flavour material) is carried out by adding various concentrations of compound Y to replicate broths containing relatively high, standard numbers of bacteria. After a certain period allowing any antibacterial activity to take place, aliquots of the bacterial cultures are diluted (usually in 10-fold steps) and dispensed onto agar plates. The plates are incubated with the expectation that each viable cell should produce a visible colony. The numbers of colonies are multiplied to take account of the dilution, to establish the number of viable cells in the broths. Once again, the broths containing compound Y are compared with an untreated control broth. The minimum concentration of compound Y which causes a reduction in the viable number of bacteria is the minimum bactericidal concentration (MBC). MBC can also be expressed in terms of the MBC required to produce a certain degree of killing (for example, a $3 \log_{10}$ reduction in count, equivalent to a 99.9% kill). Still further, the MBC can be expressed in kinetic terms - the time of exposure to an agent required for a given MBC effect.

A further possibility is that the process of inhibition could be sub-lethal (or sub-MIC), whereby the flavour materials interfere with the metabolic process, but typically do not inhibit bacterial growth.

Three modes of inhibiting the production of lactic acid are possible. In the first mode, the flavour materials (or flavour compositions) may act by direct (overt antimicrobial) killing of oral cavity bacteria, e.g. by more than 10-fold; in the second mode, they may inhibit acid generation whilst maintaining a microbial cell viability of at least 70%; in the third mode, they may inhibit acid generation at a concentration below the minimum inhibitory concentration (MIC), determined as described in Example 2 below. The third mode is preferred, since this provides anti-caries benefits, whilst leaving the natural oral cavity microflora undisturbed. Thus, preferably, the bacterial production of acid can be reduced or eliminated without significantly disturbing the oral cavity's natural microflora. This may be achieved by inhibiting the bacteria responsible for the production of acid, in particular *Streptococcus mutans*, at a concentration below the MIC.

In an even further aspect the present invention provides use of one or more of the flavour materials of the flavour composition of the invention, for the purpose of reducing and/or preventing dental caries.

The flavour composition typically also includes other flavour ingredients (which may be selected from the 400-500 or so flavour materials that are in current use when formulating flavour compositions) chosen to give desired overall flavour characteristics to the composition.

The flavour composition of the invention can be readily made by simply mixing the specified ingredients, as is well known to those skilled in the art.

The flavour compositions of the invention find application in a wide range of consumer products, particularly oral care products such as toothpastes, mouthwashes, chewing gum (where the term "chewing gum" is intended also to encompass bubble gum), confectionery, dental floss, dissolvable mouth films, breath sprays and breath freshening tablets.

The present invention also includes within its scope consumer products, particularly oral care products, including a flavour composition in accordance with the invention.

The consumer products, particularly oral care products, which include a flavour composition in accordance with the invention may be formulated in a conventional manner as is well known to those skilled in the art. For example, a toothpaste formulation will typically include from 0.3% to 2.0% by weight, preferably from 0.5% to 1.5% by weight, and more preferably from 0.8% to 1.2% by weight, of the flavour composition. A mouthwash will typically contain the flavour composition in an amount in the range 0.05% to 2.0% by weight, preferably from 0.1% to 1.0% by weight, and more preferably from 0.15% to 0.5% by weight. For a chewing gum, the composition of the invention may be present in an amount in the range 0.5% to 3.5% by weight, preferably from 0.75% to 2.0% by weight, and more preferably from 1.0% to 1.75% by weight.

A consumer product may conveniently also include ingredients such as fluoride, zinc salts, pyrophosphates etc, known to have an effect in reducing and/or preventing dental caries. These ingredients can be present in lower amounts than is typically conventional.

In an even further aspect, the present invention provides a consumer product comprising a flavour composition in accordance with the invention; and a fluoride-ion providing compound.

The fluoride-ion providing compound is capable of releasing fluoride ions or fluoride-containing ions in water. Suitable fluoride-ion providing compounds include, for example, sodium fluoride, potassium fluoride, ammonium fluoride, cuprous fluoride, zinc fluoride, stannic fluoride, stannous fluoride, barium fluoride, sodium fluorosilicate, ammonium fluorosilicate, sodium fluorozirconate, sodium monofluorophosphate, aluminium mono-and difluorophosphate and fluorinated sodium calcium pyrophosphate.

The invention also covers a consumer product comprising a flavour composition in accordance with the invention and xylitol.

It has been found by the present inventors that the combination of xylitol and a flavour composition of the invention can produce a synergistic effect, with the xylitol and flavour composition giving a greater combined effect in reducing the production of acid by *Streptococcus mutans* than xylitol and flavour composition alone. The potential synergy between xylitol and a flavour composition in accordance with the invention can be investigated using the method described in Example 4 below.

The invention will be illustrated by the following examples.

Example 1(a): Microtitre Total Acid Inhibition (TAI) Test

The following method was used to determine the efficacy of a flavour material or flavour composition at inhibiting acid production by the micro-organism *Streptococcus mutans*:

250ml of PM broth (containing: peptone, 2% w/v; tryptone, 1% w/v; yeast extract, 1% w/v; KCl, 0.25% w/v; of approximately pH 7) was charged to a Duran bottle bunged with a breathable stopper and inoculated with the test strain *Streptococcus mutans* R9, deposited under the Budapest Treaty with National Collections of Industrial, Food and Marine Bacteria Limited, 23 St Machar Drive, Aberdeen, AB24 3RY, Scotland, UK on 22nd January 2004 and given accession number NCIMB 41209 (may also be obtained from Prof. Philip Marsh, Centre for Applied Microbiology and Research, Salisbury, Wiltshire, SP4 0JG, UK). The bacterial culture was incubated anaerobically at 37°C for 48 hours. The optical density of the culture at 540 nm (OD₅₄₀) was measured and adjusted (if required) to between 0.2 and 0.3, by diluting with fresh PM broth to give a stock inoculum culture.

Acid indicator broth (AIB) was prepared by adding 4% (w/v) glucose and 0.8% (v/v) of a Bromocresol Purple stock solution (stock solution contains 16g Bromocresol Purple in 1000ml ethanol) to 0.3% (w/v) TSB broth (tryptone soya broth, available from Oxoid, Basingstoke, UK). The resulting AIB was sterilised by aseptically passing the solution through a 0.22μm filter into a sterile bottle.

Stock solutions of flavour material(s) or flavour composition(s) (flavour(s)) were made to 10,000ppm by adding 50mg of neat flavour material/flavour to 5ml of AIB, and vigorously mixing the mixture by vortex. Each row of a standard, 96-well plastic microtitre plate (labelled A-H) was allocated to one sample, thus eight samples per plate. Row H contained only Schaedler broth for use as a bacterial control to indicate the degree of turbidity resulting from bacterial growth in the absence of any test material. Aseptically, 200μl of the initial dilution of flavour material/flavour was transferred to the 1st and 7th well of the appropriate row. All other test wells were filled with 100μl of sterile Schaedler broth using an 8-channel micro-pipette. The contents of each of the wells in column 1 were mixed by sucking samples up and down in pipette tips, before 100μl was transferred

to column 2. The same sterile pipette tips were used to transfer 100 μ l of each well in column 7, into the appropriate well in column 8. This set of eight tips was then discarded into disinfectant solution. Using eight fresh, sterile tips the process was repeated by transferring 100 μ l from column 2 into column 3 (and 8 into 9). The process was continued until all wells in columns 6 and 12 contained 200 μ l. After mixing, 100 μ l was discarded from each of the wells in columns 6 and 12 to waste. Finally, 100 μ l of the bacterial stock inoculum culture was added to all wells (except the control, no bacteria wells in row H), thus giving a final volume of 200 μ l in each well. The final concentration of ingredients was thus 5,000ppm in columns 1 and 7; 2,500ppm in columns 2 and 8; and so forth so that the final concentration of ingredients in columns 6 and 12 was 156ppm.

The plates were incubated anaerobically (80% N₂, 10% H₂, 10% CO₂) for 24 hours at 37°C. Following incubation, the plates were read by eye. If the wells of bacteria/broth remained purple then the flavour material/flavour had successfully inhibited lactic acid production by *S. mutans*. If the wells of bacteria/broth appeared yellow, then *S. mutans* had metabolised glucose to lactic acid and the flavour material/flavour had not inhibited acid production.

Results were recorded as the lowest concentration at which the flavour material/flavour inhibited acid production.

Example 1(b): Bottle Total Acid Inhibition (TAI) Test

250ml of PM broth (of formulation as described in Example 1(a) above) was charged to a Duran bottle bunged with a breathable stopper and inoculated with a loopful of *Streptococcus mutans* R9 (as above). The bacterial culture was then incubated anaerobically for 2-3 days at 37°C, followed by centrifugation at 3600rpm for 10 minutes. The supernatant was decanted to waste. The pellets remaining were resuspended in 12ml of 0.1% peptone and the optical density at 540nm (OD₅₄₀) measured and adjusted (if required) by diluting with fresh PM broth to between 0.2 and 0.3 to give a stock inoculum culture.

Broth was prepared by adding 4% (w/v) glucose to 0.3% (w/v) TSB broth (GTSB). The broth was sterilised by aseptically passing the solution through a 0.22 μ m filter into a sterile bottle.

Control incubations were prepared by adding 2.5ml of the stock inoculum culture (adjusted to an OD₅₄₀ of 0.2-0.3) to 2.5ml of GTSB containing 4% (w/v) glucose in 0.3% (w/v) TSB.

A test flavour material/flavour was diluted in the GTSB to give a stock solution of flavour material/flavour with a final concentration of 25,000ppm (250mg flavour material/flavour in 10ml of GTSB).

Flavour material/flavour incubations were prepared by adding 2.5ml of the stock inoculum culture to 2.45ml of GTSB, and 50 μ l of flavour material/flavour stock solution. Thus, flavour materials/flavours were tested at a final concentration of 250ppm, for their efficacy in inhibiting acid production from 2% glucose.

The mixtures were then incubated anaerobically.

After anaerobic incubation of the resulting mixtures for 18-24 hours, the pH of the suspensions was measured using a 476530M combination pH electrode (Mettler Toledo, 64 Boston Road, Beaumont Leys, Leicester, LE4 1AW), calibrated using pH 4 and pH 7 buffers. Results were recorded as the difference in pH change between broths containing flavour material/flavour and an untreated control.

Example 2: Minimum Inhibitory Concentration (MIC)

The minimum inhibitory concentration of a flavour material or flavour composition (flavour) was determined by the following method.

A culture of the test strain *Streptococcus mutans* R9, deposited under the Budapest Treaty with National Collections of Industrial, Food and Marine Bacteria Limited, 23 St Machar Drive, Aberdeen, AB24 3RY, Scotland, UK on 22nd January 2004 and given accession number NCIMB 41209 (may also be obtained from Prof. Philip Marsh, Centre for Applied Microbiology and Research, Salisbury, Wiltshire, SP4 0JG, UK) was grown in 250ml of PM broth (containing: peptone, 2% w/v; tryptone, 1% w/v; yeast extract, 1% w/v; KC1, 0.25% w/v; of approximately pH 7), anaerobically at 37°C for 48 hours. The optical density of the culture at 540 nm (OD₅₄₀) was measured and adjusted to 0.2-0.3 by diluting with fresh PM broth. The culture was then diluted in Schaedler broth (Oxoid, Basingstoke, UK) in a ratio of 1 part culture to 25 parts broth to give a stock inoculum culture.

Flavour or flavour materials were diluted in sterile Schaedler broth to yield a 10,000 ppm stock solution, and the mixture vigorously mixed by vortex. Each row of a standard, 96-well plastic microtitre plate (labelled A-H) was allocated to one sample, thus eight samples per plate. Row H contained only Schaedler broth for use as a bacterial control to indicate the degree of turbidity resulting from bacterial growth in the absence of any test material. Aseptically, 200µl of the initial dilution of flavour/flavour material was transferred to the 1st and 7th well of the appropriate row. All other test wells were filled with 100µl of sterile Schaedler broth using an 8-channel micro-pipette. The contents of each of the wells in column 1 were mixed by sucking samples up and down in pipette tips, before 100µl was transferred to column 2. The same sterile pipette tips were used to transfer 100µl of each well in column 7, into the appropriate well in column 8. This set of eight tips was then discarded into disinfectant solution. Using eight fresh sterile tips the process was repeated by transferring 100µl from column 2 into column 3 (and 8 into 9). The process was continued until all wells in columns 6 and 12 contained 200µl. After mixing, 100µl was discarded from each of the wells in columns 6 and 12 to waste. Finally, 100µl of the pre-diluted stock inoculum culture was added to all wells (except the control, no bacteria wells in row H), thus giving a final volume of 200µl in each well.

A blank plate was prepared for each set of eight samples by repeating the process described above, except that 100 μ l of Schaedler broth was added instead of bacterial culture. This plate was used as the control plate against which the test plate(s) could be read.

Test and control plates were sealed using autoclave tape and incubated for 48 hours anaerobically at 37°C.

A microtitre plate reader (Model MRX, Dynatech Laboratories) was preset to gently agitate the plates and mix the contents. The absorbance at 540nm "A₅₄₀" was used as a measure of turbidity resulting from bacterial growth. The control, un-inoculated plate for each set of samples was read first, and the plate reader then programmed to use the control readings to blank all other plate readings for the inoculated plates for the same set of test materials (i.e. removing turbidity due to flavour and possible colour changes during incubation). Thus, the corrected readings generated were absorbances resulting from turbidity from bacterial growth. The MIC was taken as the concentration of flavour/flavour material required to inhibit growth so that the change in absorbance during the incubation period was <0.2 A₅₄₀.

Example 3

Flavour materials useful in a flavour composition of the invention were tested at 250ppm for their potential synergy with fluoride as described below.

250ml of PM broth (of formulation as described in Example 1(a) above) was charged to a Duran bottle bunged with a breathable stopper and inoculated with a loopful of *Streptococcus mutans* R9 (as above). The bacterial culture was then incubated anaerobically for 2-3 days at 37°C, followed by centrifugation at 3600rpm for 10 minutes. The supernatant was decanted to waste. The pellets remaining were resuspended in 12ml of 0.1% peptone and the optical density at 540nm (OD₅₄₀) measured and adjusted (if

required) by diluting with fresh PM broth to between 0.2 and 0.3 to give a stock inoculum culture.

Broth was prepared by adding 4% (w/v) glucose to 0.3% (w/v) TSB broth (GTSB). The broth was sterilised by aseptically passing the solution through a 0.22 μ m filter into a sterile bottle.

Control and flavour material incubations were prepared as described in Example 1(b) above.

A fluoride stock solution (2,500ppm F) was prepared by dissolving 0.552g NaF in 10ml of GTSB. For fluoride controls, 2.5ml of stock inoculum culture was added to 2.48ml of GTSB, and 20 μ l of fluoride stock solution.

For flavour material and fluoride incubations, 2.5ml of stock inoculum culture was added to 2.43ml of GTSB, 50 μ l of flavour material stock solution and 20 μ l of fluoride stock solution.

The mixtures were incubated anaerobically.

After anaerobic incubation of the resulting mixtures for 18-24 hours, the pH of the suspensions was measured using a 476530M combination pH electrode (Mettler Toledo, 64 Boston Road, Beaumont Leys, Leicester, LE4 1AW), calibrated using pH 4 and pH 7 buffers. Results were recorded as the difference in pH change between broths.

If the pH of the incubated broth containing flavour material and fluoride was higher than that measured for incubated broths containing either flavour material or fluoride, then this was considered to indicate that there had been a synergistic effect between the flavour material and fluoride in reducing the acid production of *Streptococcus mutans*.

The results are presented below, where

+ + + + = Inhibition of acid production by an additional 0.75 pH units or more (i.e. > 0.75 pH units in addition to the effect of fluoride or flavour material alone);
 + + + = Inhibition of acid production by an additional 0.50-0.74 pH units;
 + + = Inhibition of acid production by an additional 0.25-0.49 pH units; and
 + = Inhibition of acid production by an additional 0.01-0.24 pH units.

Flavour Material (at 250ppm)	Synergy with Fluoride
Alcohol C10 (Decanol)	+ + + +
Aldehyde C10 (Decanal)	+ +
Anethole Synthetic	+ + + +
Basil Comores	+ + + +
Cinnamic Aldehyde Extra	++
Citral Natural	+
Origanum	++
Peppermint Arvensis Terpeneless ACF 153	+
Peppermint Chinese Triple Rectified (Quest)	+ + / + + +
Clove Bud Rectified Extra	++
Ginger	+
Peppermint American Willamette Natural	+ + + +
Peppermint Indian Rectified	+
Tea Tree Pure	+
Thymol	+
Cardamom English Distilled	+ + + +
Damascone F	+ + +
Ionone Alpha	+ + / + + +
Tarragon	+

Example 4

Xylitol is a sugar substitute that has been used in many products as a non-cariogenic sweetener. The potential synergy between xylitol and a flavour material useful in the composition of the invention was investigated using a glucose/xylitol broth by the following method. Flavour materials were tested at 250ppm unless otherwise stated.

250ml of PM broth (of formulation as described in Example 1(a) above) was charged to a Duran bottle bunged with a breathable stopper and inoculated with a loopful of *Streptococcus mutans* R9 (as above). The bacterial culture was then incubated anaerobically for 2-3 days at 37°C, followed by centrifugation at 3600rpm for 10 minutes. The supernatant was decanted to waste. The pellets remaining were resuspended in 12ml of 0.1% peptone and the optical density at 540nm (OD₅₄₀) measured and adjusted (if required) by diluting with fresh PM broth to between 0.2 and 0.3 to give a stock inoculum culture.

Broth was prepared by adding 4% (w/v) glucose to 0.3% (w/v) TSB broth (GTSB). The broth was sterilised by aseptically passing the solution through a 0.22μm filter into a sterile bottle.

Control and flavour material incubations were prepared as described in Example 1(b) above. Xylitol synergy was investigated by adding 2.5ml of stock inoculum culture to 2.5ml of GTSB supplemented with 4% (w/v) xylitol. An additional control was also prepared with 2.5ml of stock inoculum culture added to 2.5ml of a 4% (w/v) xylitol solution in 0.3% TSB (no glucose).

These mixtures were incubated anaerobically.

After anaerobic incubation of the resulting mixtures for 18-24 hours, the pH of the suspensions was measured using a 476530M combination pH electrode (Mettler Toledo, 64

Boston Road, Beaumont Leys, Leicester, LE4 1AW), calibrated using pH 4 and pH 7 buffers. Results were recorded as the difference in pH change between broths.

If the pH of the incubated broth containing flavour material and xylitol was higher than that recorded for incubated broths containing either flavour material or xylitol, then this was considered to indicate that there was a synergy between the flavour material and xylitol in reducing the acid production of *Streptococcus mutans*.

The results for some flavour materials useful in a composition of the invention are presented below, where the degrees of synergy were allocated as described in Example 3.

Flavour Material (at 250ppm unless stated)	Synergy with Xylitol
Ginger	+++
Ionone Alpha (125ppm)	+++
Alcohol C9 (62.5ppm)	++
Basil Comores	++
Damascone	++
Aldehyde C9	+/-
Origanum (125ppm)	+/-
Aniseed rectified	+
Peppermint Aspen	+
Peppermint Moroccan	+

Ginger	+++
Ionone Alpha (125ppm)	+++
Alcohol C9 (62.5ppm)	++
Basil Comores	++
Damascone	++
Aldehyde C9	+/-
Origanum (125ppm)	+/-
Aniseed rectified	+
Peppermint Aspen	+
Peppermint Moroccan	+

Example 5

A flavour composition in accordance with the invention was prepared by mixing the following ingredients:

Ingredient	%w/w	Group
C9 Aldehyde (nonanal)	0.1	(b)

Anethole Synthetic	9.0	(a)
Cis 3 Hexenyl Butyrate	2.0	
Menthol Laevo	45.0	
Orange Oil	4.5	
Origanum	0.9	(a)
Peppermint American Yakima Rectified	31.5	(b)
Peppermint Arvensis Terpeneless	7.0	(b)
	<hr/>	
Total	100	

Example 6

A flavour composition in accordance with the invention was prepared by mixing the following ingredients:

Ingredient	% w/w	Group
Alcohol C9 (nonanol)	0.15	(a)
Anethole Synthetic	8.50	(a)
Cinnamic Aldehyde	2.25	(a)
Citral	6.60	(b)
Menthol laevo	42.50	
Orange Oil	4.25	
Peppermint American Yakima Rectified	29.75	(b)
Peppermint Arvensis Terpeneless	6.00	(b)
	<hr/>	
Total	100	

Example 7

A flavour composition in accordance with the invention was prepared by mixing the following ingredients:

Ingredient	%w/w	Group
Anethole Synthetic	7.0	(a)
Clove Bud Oil Rectified	6.0	(b)
Menthol laevo	35.0	
Orange Oil	3.5	
Peppermint American Yakima Rectified	36.4	(b)
Peppermint Chinese Triple Rectified	12.0	(b)
Origanum	0.1	(a)
	—	
Total	100	

Example 8: Formulations

Any one of the flavour compositions of Examples 5 to 7 above may be included in the following toothpaste, mouthwash, or chewing gum formulations, which are prepared according to conventional methods known to those skilled in the art:

Chalk Toothpaste

Material	%w/w
Glycerine	20.0
Distilled Water	35.3
Calcium Carbonate (Sturcal H)	40.0
Sodium Carrageenate (Viscarin)	2.00
Sodium Saccharin	0.20
Sodium Lauryl Sulphate (Empicol LZPV/C)	1.50
Flavour Composition	1.00
	—
Total	100.00

where Sturcal H, Viscarin and Empicol LZPV/C are all Trade Marks.

Opacified Silica Toothpaste

Material	%w/w
Sorbitol 70% syrup	50.0
Distilled Water	23.6
Sodium Monofluorophosphate	0.80
Trisodium Phosphate 12H ₂ O	0.10
Sodium Saccharin	0.20
Precipitated Silica (AC 30)	8.00
Precipitated Silica (TC 15)	8.00
Sodium Carboxy Methyl Cellulose (9M31XF)	0.80
Titanium Dioxide (Tiona)	1.00
Sodium Lauryl Sulphate (Empicol LZPV/C)	1.50
Polyethylene Glycol 1500	5.00
Flavour Composition	1.00
Total	100.00

Where Tiona and Empicol LZPV/C are Trade Marks.

Ready-to-Use MouthwashMixture A - Alcohol Phase

	%w/w
Ethanol 96%, Double Rectified	12.000
PEG 40 Hydrogenated Castor Oil (Cremophor RH40)	0.250
Flavour Composition	0.200

Mixture B - Aqueous Phase

	%w/w
Sorbitol 70% syrup	12.000

Saccharin 25% solution	0.200
Cetyl Pyridinium Chloride	0.025
Distilled Water	75.325

Where Cremophor RH40 is a Trade Mark.

The alcohol phase (mixture A) and aqueous phase (mixture B) were prepared separately and then combined to give the mouthwash.

Chewing Gum

Material	%w/w
Gum Base Balear T	28.0
Sorbitol Powder	52.9
Sorbitol Syrup	6.0
Xylitol	6.0
Glycerol 98 %	5.0
Aspartame	0.05
Acesulfame K	0.05
Flavour Composition	2.0

where Balear T and Acesulfame K are Trade Marks.

CLAIMS

1. A flavour composition comprising at least two flavour materials selected from the following Group (a) materials: decanol, nonanol, decanal, anethole synthetic, cardamom oil, cinnamic aldehyde, ionone alpha, origanum, tarragon, thymol; and at least one flavour material selected from the following Group (b) materials: nonanal, Aniseed rectified, basil oil, camomile oil, citral, clove bud oil, Damascone F, ginger, Tea Tree Pure, peppermint oil of natural origin.
2. A flavour composition according to claim 1, wherein the flavour composition comprises at least three flavour materials from Group (a).
3. A flavour composition according to claim 1 or 2, wherein the flavour composition comprises at least two flavour materials from Group (b).
4. A flavour composition according to any one of the preceding claims, wherein the flavour composition comprises at least 3% by weight, preferably at least 6% by weight and more preferably at least 10% by weight, of flavour materials from Group (a).
5. A flavour composition according to any one of the preceding claims, wherein the flavour composition comprises at least 3% by weight, preferably at least 10% by weight and more preferably at least 25% by weight, of flavour materials from Group (b).
6. A consumer product comprising a flavour composition according to any one of claims 1 to 5.
7. A consumer product according to claim 6, wherein the consumer product further comprises a fluoride-ion providing compound.

8. A consumer product according to claim 6 or 7, wherein the consumer product is an oral care product.
9. A method for reducing or preventing acid-producing bacteria by introducing in the oral cavity a flavour composition according to any one of claims 1 to 5.
10. Use of one or more of the following flavour materials decanol, nonanol, decanal, anethole synthetic, cardamom oil, cinnamic aldehyde, ionone alpha, origanum, tarragon, thymol, nonanal, Aniseed rectified, basil oil, camomile oil, citral, clove bud oil, Damascone F, ginger, Tea Tree Pure, peppermint oil of natural origin, for the purpose of reducing and/or preventing dental caries.

INTERNATIONAL SEARCH REPORT

PCT/GB2004/000515

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 A61K7/16

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EP0-Internal, WPI Data, PAJ, CHEM ABS Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	GB 1 591 342 A (INT FLAVORS & FRAGRANCES INC) 17 June 1981 (1981-06-17) claims 1,10,11; example XXXVIII -----	1-6
X	US 4 689 214 A (NILES HOLLANDRA P ET AL) 25 August 1987 (1987-08-25) column 8, line 10, paragraph 42; claims 1,4,11,12 -----	1,4-9
X	EP 0 950 652 A (KURARAY CO) 20 October 1999 (1999-10-20) page 11, line 1 - line 15 -----	1,6
X	US 4 335 002 A (MUSSINAN CYNTHIA J ET AL) 15 June 1982 (1982-06-15) column 19, line 1 - line 10; example XVII ----- -/-	1,3,5-9

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- "&" document member of the same patent family

Date of the actual completion of the international search

21 May 2004

Date of mailing of the International search report

28.06.2004

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Voyiazoglou, D

INTERNATIONAL SEARCH REPORT

PCT/GB2004/000515

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 4 226 892 A (DEMOLE EDOUARD ET AL) 7 October 1980 (1980-10-07) example 58 -----	1,3,6
X	US 3 966 819 A (SCHULTE-ELTE KARL-HEINRICH ET AL) 29 June 1976 (1976-06-29) example 4 -----	1,6
X	US 3 917 613 A (GUTH GERARD ET AL) 4 November 1975 (1975-11-04) example XVIII -----	1,6,8,9
X	PATENT ABSTRACTS OF JAPAN vol. 0120, no. 96 (C-484), 29 March 1988 (1988-03-29) & JP 62 230712 A (NICHIBAI BOEKI KK), 9 October 1987 (1987-10-09) abstract -----	10

INTERNATIONAL SEARCH REPORT

PCT/GB2004/000515

Box II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. Claims Nos.: 9, 10 because they relate to subject matter not required to be searched by this Authority, namely:
Although claim 9,10 are directed to a method of treatment of the human/animal body, the search has been carried out and based on the alleged effects of the compound/composition.
2. Claims Nos.: because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
3. Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this International application, as follows:

1. As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
4. No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

The additional search fees were accompanied by the applicant's protest.

No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

PCT/GB2004/000515

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
GB 1591342	A 17-06-1981	US 4062894 A		13-12-1977
		US 4021488 A		03-05-1977
		US 4068012 A		10-01-1978
		DE 2700688 A1		21-07-1977
		DE 2759425 B1		17-07-1980
		FR 2338240 A1		12-08-1977
		JP 52106809 A		07-09-1977
		JP 56138136 A		28-10-1981
		NL 7700367 A ,C		19-07-1977
		NL 8304146 A		02-04-1984
		NL 8304147 A		02-04-1984
		US 4081481 A		28-03-1978
		US 4107093 A		15-08-1978
		US 4103036 A		25-07-1978
		US 4122120 A		24-10-1978
		US 4102928 A		25-07-1978
		US 4081479 A		28-03-1978
		US 4064281 A		20-12-1977
		US 4073751 A		14-02-1978
		US 4117015 A		26-09-1978
		US 4175569 A		27-11-1979
		US 4119576 A		10-10-1978
		US 4147727 A		03-04-1979
		US 4081483 A		28-03-1978
		US 4122122 A		24-10-1978
		US 4097416 A		27-06-1978
US 4689214	A 25-08-1987	AT 389445 B		11-12-1989
		AT 87786 A		15-05-1989
		AU 587241 B2		10-08-1989
		AU 5537986 A		23-10-1986
		BE 904604 A1		16-10-1986
		BR 8601690 A		16-12-1986
		CA 1283055 C		16-04-1991
		CH 674309 A5		31-05-1990
		DE 3610179 A1		23-10-1986
		DK 173586 A		17-10-1986
		ES 8706435 A1		16-09-1987
		FI 861281 A ,B,		17-10-1986
		FR 2587209 A1		20-03-1987
		GB 2173701 A ,B		22-10-1986
		GR 860981 A1		22-12-1986
		HK 18492 A		13-03-1992
		IN 165260 A1		09-09-1989
		IT 1214408 B		18-01-1990
		MX 164084 B		13-07-1992
		NL 8600957 A		17-11-1986
		NO 861469 A ,B,		17-10-1986
		NZ 215697 A		27-01-1989
		PH 22873 A		19-01-1989
		PT 82387 A ,B		01-05-1986
		SE 467609 B		17-08-1992
		SE 8601655 A		17-10-1986
		SG 4992 G		20-03-1992
		JP 2037502 C		28-03-1996
		JP 7076167 B		16-08-1995
		JP 61263912 A		21-11-1986
		ZA 8602386 A		25-11-1987

INTERNATIONAL SEARCH REPORT

Information on patent family members

PCT/GB2004/000515

Patent document cited in search report	Publication date		Patent family member(s)	Publication date
EP 0950652	A	20-10-1999	JP 11292810 A AT 220057 T CN 1233611 A ,B DE 69901979 D1 DE 69901979 T2 EP 0950652 A1 US 6159928 A JP 2000226357 A	26-10-1999 15-07-2002 03-11-1999 08-08-2002 27-02-2003 20-10-1999 12-12-2000 15-08-2000
US 4335002	A	15-06-1982	US 4241098 A US 4267066 A US 4312766 A US 4319586 A US 4280933 A	23-12-1980 12-05-1981 26-01-1982 16-03-1982 28-07-1981
US 4226892	A	07-10-1980	CH 528470 A CH 529709 A CH 520767 A CH 520479 A CH 498795 A CH 536834 A CH 539682 A DD 96077 A6 DE 2022216 A1 DE 2065322 A1 DE 2065323 A1 DE 2065324 A1 FR 2042452 A5 GB 1305621 A IT 1047883 B JP 55003328 B NL 7006649 A ,B, US 3975310 A US 3928456 A US 3931326 A US 4187863 A CH 536603 A BE 750049 A4 CH 528225 A CH 521099 A CH 513096 A CH 513097 A CH 513094 A CH 521298 A CH 513098 A CH 537452 A CH 537352 A CH 536604 A BE 723610 A CH 524560 A CH 505773 A CH 509399 A CH 524320 A DE 1807568 A1 DE 1817884 A1 ES 359994 A1 ES 377482 A1 FR 1591031 A	30-09-1972 31-10-1972 31-03-1972 31-03-1972 15-11-1970 15-05-1973 31-07-1973 12-03-1973 19-11-1970 29-03-1973 22-03-1973 29-03-1973 12-02-1971 07-02-1973 20-10-1980 24-01-1980 10-11-1970 17-08-1976 23-12-1975 06-01-1976 12-02-1980 15-05-1973 06-11-1970 30-09-1972 15-04-1972 30-09-1971 30-09-1971 30-09-1971 15-04-1972 30-09-1971 13-07-1973 31-05-1973 15-05-1973 08-05-1969 30-06-1972 15-04-1971 30-06-1971 30-06-1972 19-06-1969 29-03-1973 01-10-1970 01-07-1972 20-04-1970

INTERNATIONAL SEARCH REPORT

PCT/GB2004/000515

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
US 4226892	A	GB 1240309 A		21-07-1971
		GB 1240310 A		21-07-1971
		IT 1034009 B		10-09-1979
		JP 48029215 B		08-09-1973
		JP 48029216 B		08-09-1973
		JP 48031105 B		26-09-1973
		JP 48029217 B		08-09-1973
<hr/>				
US 3966819	A 29-06-1976	CH 585522 A5		15-03-1977
		DE 2440025 A1		13-03-1975
		FR 2242360 A1		28-03-1975
		GB 1418620 A		24-12-1975
		JP 974008 C		19-10-1979
		JP 50058035 A		20-05-1975
		JP 54006615 B		30-03-1979
		NL 7411449 A ,B,		07-03-1975
		US 3965189 A		22-06-1976
<hr/>				
US 3917613	A 04-11-1975	GB 1436329 A		19-05-1976
		AT 329767 B		25-05-1976
		AT 691773 A		15-08-1975
		AU 463818 B2		07-08-1975
		AU 5881873 A		06-02-1975
		BE 803317 A1		07-02-1974
		CA 1030965 A1		09-05-1978
		CH 604559 A5		15-09-1978
		DE 2339661 A1		21-02-1974
		FR 2195630 A1		08-03-1974
		IT 1048413 B		20-11-1980
		JP 49055880 A		30-05-1974
		JP 53027780 B		10-08-1978
		NL 7310916 A ,B,		11-02-1974
		NO 138014 B		06-03-1978
		ZA 7305315 A		26-03-1975
<hr/>				
JP 62230712	A 09-10-1987	NONE		
<hr/>				